

RT-QuIC buffers recipe

- **2M NaCl**

5.845g of NaCl in 50 ml of mQ water

- **5X PBS**

Dilute 1:1 10X PBS (see recipe below) in mQ water
(25 ml 10X PBS + 25 ml mQ water)

- **0.1% SDS/PBS**

0.05g SDS + 10 ml 5X PBS + 40mL mQ water

- **100mM EDTA**

Dilute 1:5 500 mM EDTA (see recipe below) in mQ water

$M=n/VL$
$g=n*MW$
$n=g/MW$
$g= M*VL*MW$

Once ready, all the buffers must be filtered in a biosafety hood with [0.22µm Argos Tech syringe filters](#) (to keep them sterile and clean). Keep the solutions at room temperature.

- **10X PBS (pH 6.9)**

75.9 g/l NaCl

13.8 g/l $NaH_2PO_4 \cdot H_2O$ (Sodium phosphate, monobasic monohydrate)

26.8 g/l $Na_2HPO_4 \cdot 7H_2O$ (Sodium phosphate, dibasic heptahydrate)

Once all the powders are dissolved the pH to 6.9 and filter with 0.22µm filters, in the hood.

pH should increase by 0.5 unit when diluted to 1X so it will be 7.4 (and 130 mM NaCl)

- **500mM EDTA (pH 8.0)**

146.12 g/l EDTA powder (weigh 452.23 g if using tetrasodium EDTA tetrahydrate)

Dissolve powder into 900 ml of mQ water. EDTA won't go into solution if the pH is acidic, you need to add 10M NaOH (or ~8 pellets of NaOH) in order to increase the pH to 8. Then bring to 1 l volume and filter the solution with 0.22µm filters.

Pro tips: use Falcon tubes, other brands don't seal tightly enough.